
The evolution of nematode sex determination: *C. elegans* as a reference point for comparative biology*

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Abstract

Sex determination was a founding topic of *C. elegans* research. After three decades of research, a complex signal transduction pathway with multiple layers of regulation has been elucidated. This pathway links karyotype to phenotype by coordinating the development of sexually dimorphic tissues. In this article, this pathway is placed in two broader contexts. The first is that of nematodes and animals in general. The important role of *C. elegans* studies in revealing the first universally conserved component of metazoan sex determination is discussed, as is the role of cooption of genes into the sex determination and dosage compensation pathways. The second context is that of a subset of more closely related species, with emphasis on other members of the genus *Caenorhabditis*. Studies reviewed here have determined the gene-level conservation of the known pathway and the relative rates of molecular evolution in conserved components, and made substantial progress in the manipulation of gene activity in non-*elegans* species. Special attention is paid to the origins of hermaphroditism, which evolved from gonochorism through germline-specific changes in sex determination. Recent studies suggest that the most rapidly evolving aspects of sex determination are germline functions related to evolutionary shifts in mating systems, while somatic sex determination is relatively conservative. From all of these studies, a picture emerges in which *C. elegans* utilizes an intriguing mixture of general and species-specific genes and regulatory mechanisms.

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1. Introduction

Sex determination was one of the first aspects of *C. elegans* biology to be systematically characterized with genetic analysis (Hodgkin and Brenner, 1977), and the depth of this analysis has made it a major topic in developmental biology. It is both fair and interesting to ask how general the *C. elegans* model is likely to be, both for other nematodes as well as for animals in general. As nematodes employ many different reproductive strategies, often related to parasitic life histories (Anderson, 2000; Chitwood and Chitwood, 1950), one might expect similarly variable sex determination mechanisms. However, comparative studies of sex determination based on the *C. elegans* model have revealed both rapidly evolving and surprisingly well conserved features. This mixture of old and new (or slow and fast), along with its obvious relevance to evolution, ecology, and applied fields like agricultural and medical parasitology, has made the evolution of nematode sex determination an increasingly active research area.

2. Genetic and environmental sex determination in nematodes

In 1949, Nigon demonstrated that *C. elegans* used an XX/XO sex chromosome system, and the importance of the X:autosome ratio was clarified by Madl & Herman (1979). The XX/XO system is widespread across the Nematoda, and may be ancestral (Bull, 1981; Bull, 1983). However, some groups have apparently abandoned genetic sex determination (GSD) in favor of environmental sex determination (ESD).

The best characterized example of nematode ESD is in the arthropod parasitic Mermithidae. In lightly infected hosts, exclusively female worms are born, while in heavily parasitized hosts only males are produced. At intermediate parasite densities mixed populations are produced (Christie, 1929; Harlos et al., 1980). Culture of mermithids in axenic liquid culture also produces only females, suggesting that the concentration of a limiting host factor may be an important cue for male development (Petersen, 1985). Whether the environmental factors that dictate sex impinge upon a signal transduction pathway resembling that mediating *C. elegans* sex determination is unknown, but just such a connection between nutrition and sex determination has recently been found in *C. elegans* itself (Prahlad et al., 2003). Among the small number of nematode fossil specimens that exist are mermithids emerging from their insect hosts, preserved in 40 million year old Baltic amber (Poinar, 2002). Their striking similarity to extant insect-parasitic species (Figure 1) suggests a minimum age for the origin of ESD in this group, although it is possible that ancient mermithids used GSD.

ESD also occurs in the vertebrate parasite *Strongyloides*, which like *C. elegans* is in the order Rhabditida. As with most parasitic nematodes, a dauer-like alternative L3 form serves as the infective stage. Once inside a host, infective worms all mature as females and reproduce by mitotic parthenogenesis (Anderson, 2000). Parthenogenetic progeny that remain in the host develop into apomictic females, like their mothers. However, when L1 larvae are passed from the host in feces, they can either develop into infective L3 female larvae (the homogonic path) or initiate a free-living male/female generation (the heterogonic phase) that is similar to *Caenorhabditis*. Heterogonic development produces males and females from genetically identical L1 larvae, indicating that sex is either stochastically or environmentally determined. This again resembles the sexual plasticity of *C. elegans* larvae recently described by Prahlad et al. (2003), in which a chemical cue induces specific loss of the paternal X chromosome in L1 larvae. It would be interesting to see if these phenomena share a common mechanism.

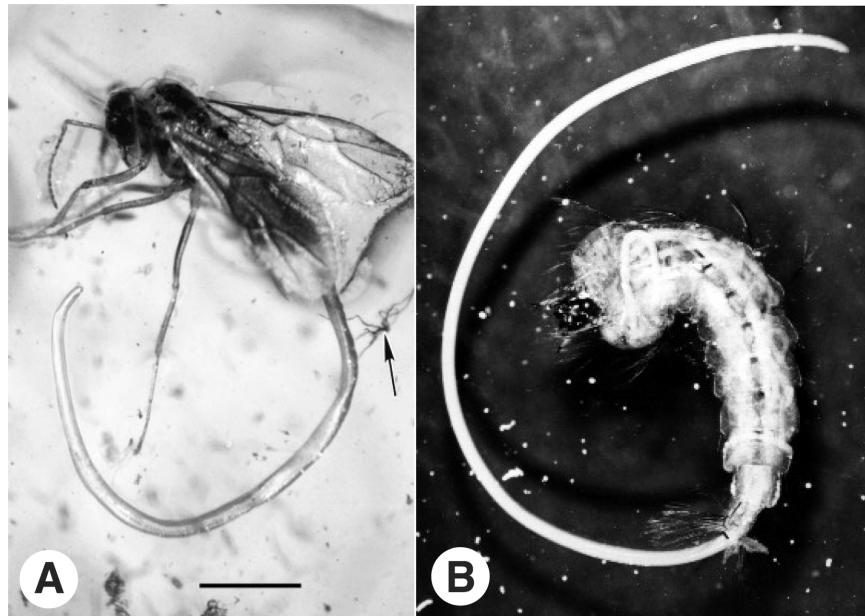


Figure 1. Ancient and extant mermithid nematodes emerging from their insect hosts. A. Juvenile worm of the genus Heydenius emerging from a winged male ant of the genus *Prenolepis*. The specimen is preserved in Baltic amber approximately 40 million years old. The scale bar represents 1.2 mm. Photograph reproduced from Poinar (2002) with permission from the author and Cambridge Univ. Press. B. Juvenile mermithid emerging from an extant mosquito larva. Photo used with permission of the Univ. of Nebraska, Lincoln Dept. of Entomology.

Commitment to heterogonic development in *Strongyloides* requires amphid neurons homologous to those that govern dauer formation in *C. elegans* (Ashton et al., 1998), indicating that the alternate reproductive modes are also environmentally controlled. Males are required for female fertility in the heterogonic phase, but apparently the male sperm serves only to activate embryogenesis. Diploidy is thought to be maintained by fusion of the post-meiotic oocyte pronucleus with an oogenic polar body (Anderson, 2000). The heterogonic phase is therefore pseudo-sexual, and presumably evolved from a truly sexual ancestor. Given that parasites often rely on genetic diversity for successful host invasion, this represents an unusual case that deserves further investigation.

3. Deep conservation and cooption in the evolution of nematode sex determination

Although *C. elegans*, *Drosophila*, and mammals (the three most-studied systems) all use GSD, the signal transduction pathways that convert the genetic difference between sex into dimorphic fates are not at all similar (Cline and Meyer, 1996). However, recently a conserved component has been discovered: the *Doublesex/mab-3* (DM) family of transcriptional regulators (Figure 2, Raymond et al., 1998; Yi and Zarkower, 1999). Although *mab-3* functions in specifying a limited number of male cell fates in *C. elegans*, the role of *Dsx* in *Drosophila* and of DM proteins in some vertebrates (Matsuda et al., 2002) indicate they can function more globally. The association of DM family member expression with sexual development in cnidarians (Miller et al., 2003) further suggests that this role evolved prior to the Cambrian, over 500 million years ago. It is reasonable to imagine that the first metazoans had sexual differentiation controlled in part by a DM progenitor, and that the diversity of extant sex determination mechanisms represents extreme divergence in different lineages. In this sense, then, sex determination itself is a homologous developmental process, even though the genes that control it are in most cases not themselves homologous.

How did the nematode system evolve? One interesting possibility is that much of it represents a highly modified form of the *hedgehog (hh)* pathway. *hh* signaling controls many key patterning and cell fate decisions in metazoan development, but is conspicuously absent from *C. elegans*, as judged by the lack of bona fide *hh* and *smoothened* homologs in the genome. However, the *TRA-2* transmembrane receptor is topologically similar to *patched*, and the pathway culminates in the regulation of a *Cubitus interruptus/Gli*-like transcription factor, *TRA-1*. Perhaps in the distant past, as nematode development became more mosaic and increasingly dependent upon cell lineage, *hh* signaling became dispensable for other aspects of development and was coopted into sex determination. Given the unusually high rates of sequence evolution exhibited by many sex determination genes, it is plausible that

some “novel” proteins in the pathway may have at one time been recognizably similar to other *hh*-related homologs. The isolation of *her-1* (Streit et al., 1999) and *tra-1* (Pires-daSilva and Sommer, 2004) homologs from *Brugia malayi* (from the order Spirurida) and *Pristionchus pacificus* (Rhabditida, Neodiplogasteridae), respectively, suggests that this putative cooption happened early in the diversification of nematodes. In the case of *Ppa-tra-1*, multiple mutant alleles provide conclusive proof of a conserved female-promoting role.

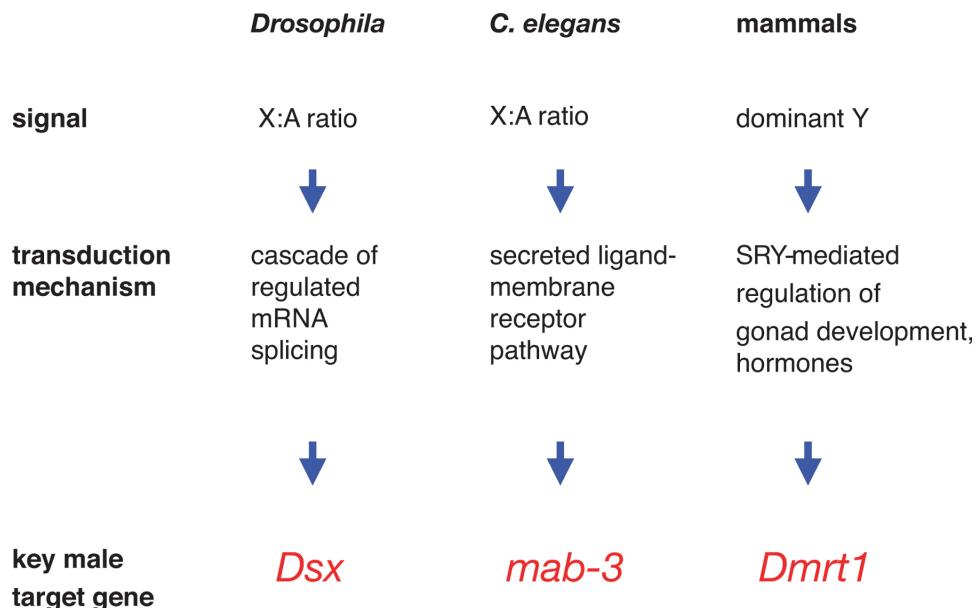


Figure 2. Variation in model system sex determination. Though mammals, *Drosophila*, and *C. elegans* all use GSD, they interpret their sex chromosome content through distinct signal transduction pathways. However, all three eventually converge on a DM family member whose expression is associated with male development. *Drosophila Dsx* is unusual for DM genes in also having an important role in female development (through a female-specific splice variant).

There are two other likely cases of cooption of genes into *Caenorhabditis* sex determination, enabled through pleiotropic function of a single gene or the divergence of gene duplicates. First, the recent structural characterization of *XOL-1* suggests that it is related to small molecule metabolic kinases (Luz et al., 2003). Whether *XOL-1* actually acts as an enzyme is not yet clear, but the fact that it is the least conserved protein in the entire sex determination pathway suggests it may not. The other case is comprised of the dosage compensation complex genes. The products of *mix-1*, *dpy-26*, *dpy-27*, and *dpy-28* are clearly related to the 13S condensin complex that mediates chromatin condensation in vertebrates (Meyer, 2000). *MIX-1* has been shown to play a direct role in both processes (Hagstrom et al., 2002), while *dpy-26*, *dpy-27* and *dpy-28* are replaced in cell division by products of the paralogs *C29E4.2* (Hagstrom and Meyer, pers. comm.), *smc-4* (Hagstrom et al., 2002), and *hcp-6* (Chan et al., 2004), respectively.

In 1995, Wilkins proposed that the many layers of negative regulation found in nematode and *Drosophila* sex determination represent the accumulation of successive upstream repressors, the so-called “bottom up” hypothesis”. The conservation of *tra-1* as a key female-promoting gene beyond the family Rhabditidae is consistent with this idea. However, if the signal transduction pathway used in *Caenorhabditis* sex determination is a diverged form of the *hedgehog* pathway, then one might predict that the entire “cassette” from *her-1* to *tra-1* came into the pathway at once, and thus will not be found in partial form in any species.

A more general test of the bottom-up idea will require examination of the more upstream components of the pathway, such as the X chromosome counting factors *fox-1* and *sex-1*, in both close and distant relatives of *C. elegans*. Although the XX/XO system seems to be widespread across the Nematoda, this does not mean that X dosage is measured by homologous genes in all species. Although clear *C. briggsae* *fox-1* and *sex-1* homologs exist, their functions remain untested. As of early 2005 an effort to sequence the *Pristionchus pacificus* genome is underway, which should soon enable a search for these genes (or their equivalents) in a second nematode family. Recently, Hodgkin (2002) demonstrated that *C. elegans* can be manipulated genetically such that any of the core sex-determining genes (all autosomal), an extrachromosomal array, temperature, or even a suppressor tRNA locus can substitute for the X:A ratio as the primary determinant of sex. This impressive feat suggests that nature also has

a large number of options at its disposal for optimization of reproductive strategy. It will be of great interest to see to what extent these options have, in fact, been used during nematode evolution.

4. Conservation of *C. elegans* pathway components: *C. briggsae* as the ruler

Homologs of *C. elegans* sex determination genes have been isolated from several *Caenorhabditis* species, including the undescribed male/female species *CB5161* (Stothard et al., 2002), the male/female *C. remanei* (Chen et al., 2001; Haag and Kimble, 2000; Haag et al., 2002), and the non-rhabditids *Pristionchus pacificus* (Pires-daSilva and Sommer, 2004) and *Brugia malayi* (Streit et al., 1999). However, the nearly complete genome sequence assembly of *C. briggsae* (Stein et al., 2003) makes it uniquely suited to assess molecular conservation across the entire *C. elegans* pathway. The focus below is therefore on *C. briggsae*, but we shall return to the results of functional studies in the other species in a later section.

Nearly all *C. elegans* sex determination genes have clear *C. briggsae* orthologs, as judged by genomic synteny, reciprocal best BLAST scores, and in a few cases comprehensive phylogenies (Nayak et al., 2005; Stothard et al., 2002). Table 1 shows the amino acid conservation between these *C. elegans* and *C. briggsae* homologs. Most, but not all, of these genes show greater divergence than the genome-wide median conservation of 80% (Stein et al., 2003). Little correlation exists between where a gene acts in the pathway and its sequence conservation. One trend, however, seems to be higher conservation in proteins that are demonstrably or likely to be pleiotropic. For example, *mag-1*, *mog-1*, *mog-4*, and *mog-5* all encode key pre-mRNA splicing factors (Belfiore et al., 2004; Li et al., 2000; Puoti and Kimble, 1999; Puoti and Kimble, 2000), and are extremely well conserved. Similarly, the RNA-binding *GLD-1* has roles in both sex determination (Jan et al., 1999; Jones et al., 1996; Jones and Schedl, 1995a) and the mitosis-meiosis decision (Crittenden et al., 2002; Francis et al., 1995; Hansen et al., 2004; Kadyk and Kimble, 1998) in the *C. elegans* germ line.

Table 1. *C. elegans*-*C. briggsae* sex determination protein sequence conservation

Gene	Role in sex determination pathway	Protein family or motifs	Amino acid identity
<i>sex-1</i>	X dosage counting element (female-promoting)	Zn-finger TF, nuclear hormone receptor (Carmi et al., 1998)	44% B
<i>fox-1</i>	X dosage counting element (female-promoting)	RRM (Hodgkin et al., 1994)	87% B
<i>xol-1</i>	integrator of X and autosomal dosage (male-promoting)	GHMP kinase-related (Luz et al., 2003)	22% (Luz et al., 2003)
<i>sdc-1</i>	X dosage compensation complex component, <i>her-1</i> transcriptional repressor (female-promoting)	Zn-finger TF (Nonet and Meyer, 1991)	39% B
<i>sdc-2</i>	X dosage compensation complex component, <i>her-1</i> transcriptional repressor (female-promoting)	novel, with coiled-coil (Dawes et al., 1999)	32% B
<i>sdc-3</i> (aka <i>dpy-29</i>)	X dosage compensation complex component, <i>her-1</i> transcriptional repressor (female-promoting)	Zn-fingers, myosin-like ATPase (Klein and Meyer, 1993)	28% G
<i>her-1</i>	secreted signal coordinating gender (male-promoting)	novel, signal peptide (Hamaoka et al., 2004; Perry et al., 1993)	57% (Streit et al., 1999)
<i>tra-2</i>	receptor for HER-1, repressor of <i>fem</i> genes (female-promoting)	7-pass integral membrane protein (Kuwabara and Kimble, 1995)	43% (Kuwabara, 1996)
<i>tra-3</i>	positive regulator of <i>TRA-2</i> processing (female promoting)	calpain-related protease (Barnes and Hodgkin, 1996)	91% B
<i>laf-1</i>	somatic repressor of <i>tra-2</i> translation (male-promoting)	noncoding RNA (Goodwin, pers. comm.)	N/A
<i>gld-1</i>	germline translational repressor of <i>tra-2</i> (male-promoting)	KH RNA-binding protein (Jones and Schedl, 1995b)	83% B

Gene	Role in sex determination pathway	Protein family or motifs	Amino acid identity
<i>fog-2</i>	germline cofactor of GLD-1 , repressor of tra-2 translation (male-promoting)	F-box (Clifford et al., 2000)	No clear ortholog
<i>mag-1</i>	germline repressor of male-promoting genes (female-promoting)	<i>magoH/mago nashi</i> exon junction complex (Li et al., 2000)	97% G
<i>fem-1</i>	cytoplasmic responder to TRA-2 activity (male-promoting)	ankyrin repeats (Spence et al., 1990)	72% B
<i>fem-2</i>	cytoplasmic responder to TRA-2 activity (male-promoting)	protein phosphatase 2C (Pilgrim et al., 1995)	63% (Hansen and Pilgrim, 1998)
<i>fem-3</i>	cytoplasmic responder to TRA-2 activity (male-promoting)	novel	38% (Haag et al., 2002)
<i>fbf-1,2</i>	germline translational repressors of <i>fem-3</i> (female-promoting)	Puf family RNA-binding protein (Zhang et al., 1997)	No clear ortholog
<i>nos-3</i>	germline cofactor of FBF-1/2 , repressor of <i>fem-3</i> translation (female-promoting)	<i>nanos</i> -related RNA-binding domain	51% B
<i>mog-1</i>	global repressor of <i>fem-3</i> translation (female-promoting)	DEAH RNA helicase (Puoti and Kimble, 1999)	92% B
<i>mog-4</i>	global repressor of <i>fem-3</i> translation (female-promoting)	DEAH RNA helicase (Puoti and Kimble, 2000)	90% B
<i>mog-5</i>	global repressor of <i>fem-3</i> translation (female-promoting)	DEAH RNA helicase (Puoti and Kimble, 2000)	92% B
<i>mog-6</i> (aka <i>cyp-4</i>)	global repressor of <i>fem-3</i> translation (female-promoting)	divergent cyclophilin (Belfiore et al., 2004)	92% B
<i>tra-1</i>	global regulator of sex-specific transcription (female-promoting)	Zn-finger TF (Zarkower and Hodgkin, 1992)	44% (de Bono and Hodgkin, 1996)
<i>fog-1</i>	promoter of spermatogenesis (male-promoting)	CPEB (Jin et al., 2001; Luitjens et al., 2000)	60% G
<i>fog-3</i>	promoter of spermatogenesis (male-promoting)	TOB family (Chen et al., 2000)	56% (Chen et al., 2001)
<i>mab-3</i>	regulator of male tail and neuron development (male-promoting)	DM domain (Raymond et al., 1998)	67% G
Genes are arranged by where they act in the pathway, with sex chromosome counting elements at the top and tissue-specific targets of TRA-1 at the bottom. Percent identities are based on published figures when available, on BLAST 2.0 alignments of predicted genes from WormBase when alignable over at least 90% of their full length (B), or on whole alignments using the Needleman & Wunsch algorithm as implemented in the Gap program of the GCG Wisconsin Package (G). Genes that have not yet been cloned, or whose products are involved in dosage compensation but not sex determination, are omitted here for simplicity. The median amino acid identity across the entire set of <i>C. elegans</i> - <i>C. briggsae</i> orthologs is 80% (Stein and others, 2003). P, based on partial sequence; TF, transcription factor; RRM, RNA recognition motif.			

Even well-conserved proteins can rapidly change their role (or lack thereof) in sex determination, as has been found in dipteran insects (reviewed by Graham et al., 2003). Evidence for similar change in nematodes is discussed below. In contrast, some proteins exhibiting rapid sequence change participate in conserved protein-protein interactions with a consistent role in sex determination. For example, the **TRA-1**/**TRA-2c** interaction is conserved in *C. briggsae*, and this interaction is species-specific (Wang and Kimble, 2001). A more extreme case of molecular coevolution exists between **TRA-2c** and **FEM-3**. The **FEM-3** binding domain of **TRA-2c** is conspicuously hyperdivergent (Haag and Kimble, 2000; Kuwabara, 1996), yet mediates a strong but species-specific interaction with the similarly divergent **FEM-3** in *C. elegans*, *C. briggsae*, and *C. remanei* (Haag et al., 2002). Why and how crucial protein-protein interactions mediating sex determination are continually reinvented at the primary sequence level are interesting questions that will require the integration of structural biology, genetics, and ecology to be fully answered.

5. Two cases of species-specific genes

Though most *C. elegans* sex determination genes have a single *C. briggsae* ortholog, there are two exceptions. *fog-2* is clearly the result of a recent, *elegans*-specific tandem duplication (Clifford et al., 2000). *FOG-2* differs little from the protein encoded by its nearest neighbor and most similar paralog, *FTR-1*, over its N-terminal 80%, but has a distinct C-terminus. This divergent region is necessary and sufficient for *GLD-1* binding, and may be derived from the ancestral domain in part via a frame-shift mutation (Nayak et al., 2005). *fog-2* likely reveals the footprints of a key event in the evolution of hermaphroditism in the *C. elegans* lineage. This will be discussed at greater length below.

The second example of species-specific sex determination genes is that of *fbf-1* and *fbf-2*. These two Puf family genes are also the product of a recent tandem duplication, and encode proteins that are 91% identical (Zhang et al., 1997). Three *C. briggsae* genes, themselves products of recent lineage-specific duplication events, are more closely related to the *fbfs* than to any other *C. briggsae* family members (Lamont et al., 2004, A. Doty, unpublished data). However, the expectation that any of these three *C. briggsae* genes perform the function of *fbf* in repressing *fem-3* is complicated by the lack of *Cb-fem-3(RNAi)* germline feminization (Haag et al., 2002). It may be that these *fbf*-related *C. briggsae* Puf proteins regulate *Cb-fem-3*, but only in the soma, or have other targets entirely.

6. Comparative functional studies

Several labs have investigated the function of homologs of *C. elegans* sex determination genes. An early study on *C. briggsae tra-1* (de Bono and Hodgkin, 1996) employed partial rescue of *C. elegans* mutants to demonstrate conservation of function. However, the discovery of RNAi has had a profound impact. Kuwabara (1996) first showed the efficacy of RNAi in a non-*elegans* species, against *Cb-tra-2*. Although rescue assays have continued to be used (Chen et al., 2001; Hansen and Pilgrim, 1998), the phenomenon of interspecies protein-protein incompatibility discussed above makes partial or failed interspecies rescue uninformative. Unfortunately, RNAi has its own problems, the chief being its frequently lower effectiveness in *C. briggsae* and *C. remanei* than in *C. elegans* (Haag and Kimble, 2000; Haag et al., 2002; Kuwabara, 1996, M. Montgomery, pers. comm.). For this reason, several laboratories have begun forward and reverse genetic screens for true sex determination mutations in *C. briggsae* (D. Pilgrim, E. Haag, and R. Ellis, unpublished data). In *Pristionchus pacificus*, both morpholino oligonucleotides and forward mutagenesis screens have recently been employed with success (Pires-daSilva and Sommer, 2004).

Despite these technical difficulties, sex determination function has already been demonstrated for numerous orthologs of key *C. elegans* genes. *Cb-her-1* is required for normal male development, and its overexpression clearly masculinizes XX *C. elegans* animals. Overexpression of *her-1* from the filarial parasite *Brugia malayi* was less successful (Streit et al., 1999). *tra-2* has conserved roles in both the soma and germ line of *C. briggsae* (Kuwabara, 1996) and *C. remanei* (Haag and Kimble, 2000), and *in vitro* and *in vivo* evidence exists for its translational repression in these species (Haag and Kimble, 2000; Jan et al., 1997). Similarly, *tra-1* performs a key female-promoting role in other species, both across *Caenorhabditis* (de Bono and Hodgkin, 1996) and in the neodiplogasterid *Pristionchus pacificus* (Pires-daSilva and Sommer, 2004).

The *fem* genes of non-*elegans* *Caenorhabditis* species have also been studied, with more surprising results. Although *fem-1*, *fem-2*, and *fem-3* perform conserved roles in somatic sex determination, RNAi data (Haag et al., 2002; Stothard et al., 2002) and two mutations (Hill et al., unpublished data) indicate that they are dispensable for hermaphrodite spermatogenesis in *C. briggsae*. In addition, *fem-1* (A. Spence, pers. comm.) and *fem-3* (Haag et al., 2002) are probably not required for at least some male spermatogenesis. Despite these differences, however, the germline-specific *fog-1* and *fog-3* have conserved roles in promoting spermatogenesis (Chen et al., 2001). The potential significance of these results is discussed further below.

7. Mating system evolution in *Caenorhabditis*

From a phylogenetic perspective, *C. elegans* is rather unusual in having a hermaphrodite sex. Its combination with males, termed androdioecy, is found in animals only in other nematodes and some crustaceans (Sassaman, 1995; Sassaman and Weeks, 1993). With the exception of *C. briggsae*, the remaining *Caenorhabditis* species are all gonochoristic (male/female). That hermaphroditic *Caenorhabditis* are derived from gonochoristic ancestors has been accepted for many years (Honda, 1925), as has the independent origin of selfing in distantly related species (Fitch, 2002). However, recent phylogenies suggest that even *C. elegans* and *C. briggsae* acquired their shared

hermaphroditism via parallel evolution (Cho et al., 2004; Kiontke et al., 2004). An independent origin of bisexual germline development may help explain discrepancies in germline gene function that have begun to emerge from comparative functional studies.

In *C. elegans*, the core somatic pathway is also required for normal germline sex determination, although its regulation differs from the soma in some details (Figure 3A). First, a direct TRA-1/TRA-2c interaction (Lum et al., 2000; Wang and Kimble, 2001) is required for reliable hermaphrodite spermatogenesis. Lum et al. (2000) suggested that this interaction, while generally female-promoting, is a necessary prerequisite for full repression of *tra-2* that allows initiation of hermaphrodite spermatogenesis. Second, the *fem* genes positively regulate spermatogenesis (Hodgkin, 1986) and germline-specific *fog-3* transcription (Chen and Ellis, 2000) in the absence of *tra-1*. Finally, *tra-1* loss-of-function mutants cannot sustain spermatogenesis, a phenotype at odds with their complete somatic masculinization (Schedl et al., 1989). This long-puzzling result may stem from both positive and negative regulation of *fog-3* by TRA-1 (Chen and Ellis, 2000).

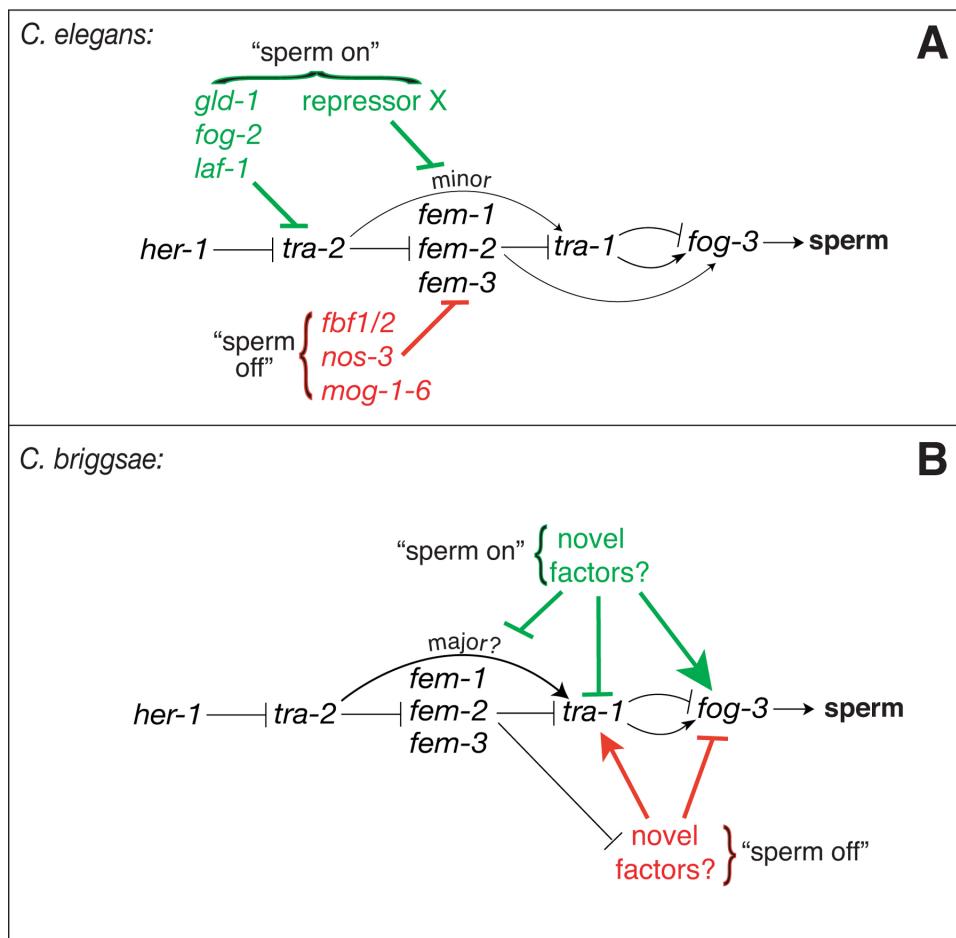


Figure 3. Models for germline sex determination in *C. elegans* and *C. briggsae*. (A) *C. elegans*, and (B) *C. briggsae*. In both panels, arrows indicate positive regulation, and crossbars indicate repressive regulation. With the exception of *fog-3*, the genes in black type are also crucial in somatic sex determination. Mechanisms promoting the initiation of hermaphrodite spermatogenesis are colored green, and those promoting the switch to oogenesis are colored red. Though it is likely that *C. briggsae* hermaphroditism is controlled by genes regulating the core pathway downstream of the *Cb-fems*, their identity is unknown. See text for full explanation.

Numerous studies have investigated the germline-specific regulation that enables hermaphrodite germ cells to change sex during development (Figure 3A). Translational repression of *tra-2* by the GLD-1/FOG-2 complex (Clifford et al., 2000; Goodwin et al., 1993; Jan et al., 1999) is thought to be the major factor allowing hermaphrodite spermatogenesis to start (“sperm on”). The switch to oogenesis depends crucially upon translational repression of *fem-3* by the FBF/NOS-3 complex (Ahringer and Kimble, 1991; Ahringer et al., 1992; Gallegos et al., 1998; Kraemer et al., 1999; Zhang et al., 1997).

How much of the regulation described above is used by *C. briggsae* hermaphrodites? There is evidence that the repression of *tra-2* by a GLD-1-like factor (Jan et al., 1999) and a TRA-1/TRA-2c direct interaction (Wang and Kimble, 2001) both occur. This is consistent with a central role for *tra-2* regulation in the initiation of *C. briggsae* hermaphrodite spermatogenesis (Figure 3B). However, *fog-2*, a key cofactor of GLD-1, is unique to *C. elegans* (see above). In addition, RNAi directed against the *C. briggsae* *gld-1* ortholog causes highly penetrant germline masculinization, not the feminization expected from *C. elegans* (Nayak et al., 2005). As discussed by Nayak et al. (2005), these latter results suggest several alternative possibilities. One is that *Cb*-GLD-1 does not translationally repress *Cb*-*tra-2*, or that it does but also has other, male-promoting targets that are more important. Alternatively, *fog-2* may qualitatively modify the function of *Ce*-GLD-1 in a conserved GLD-1/*tra-2* interaction. It is still possible, however, that the unexpected *Cb*-*gld-1*(RNAi) phenotype is due to incomplete inactivation of *Cb*-*gld-1*, as RNAi is generally less effective in *C. briggsae* (Haag et al., 2002, M. Montgomery, pers. comm.). Mutations in *Cb*-*gld-1* will be required to clarify this matter.

In addition to the evolution of the “sperm on” portion of hermaphrodite germline development, evidence is mounting that the “sperm off” component is also distinct. While the *Cb-fem* genes have conserved roles in male somatic development, none have yet been shown to be required for hermaphrodite spermatogenesis (Haag et al., 2002, A. Spence, pers. comm.; Stothard et al., 2002, Hill et al., unpublished data). Thus the “sperm off” function must target other parts of the pathway, suggested here (Figure 3B) to be variously the TRA-1/TRA-2c interaction, direct upregulation of *tra-1*, or repression of *fog-3*. *fog-3* expression is tightly associated with sperm production in both species, as well as in *C. remanei* (Chen et al., 2001). Surprisingly, *Cb-fem-2* and *Cb-fem-3* null XO mutants are neither females (Fog), as would be the case in *C. elegans*, nor somatic females with spermatogenic germ lines (Mog), as might be expected from the lack of phenotype in XX hermaphrodites. Instead, these XO mutants are also hermaphrodites (the Her phenotype; Hill et al., unpublished data). This suggests that the *Cb-fem* genes are neither required to initiate nor to curtail hermaphrodite spermatogenesis, but do act to repress the sperm-oocyte switch in XO males. Overall, it is likely the entire control of hermaphrodite spermatogenesis lies downstream of the *fem* genes in *C. briggsae* (Figure 3B).

The question of how hermaphrodites differ from their female ancestors has motivated much of the comparative work on *Caenorhabditis* sex determination. *C. briggsae* and *C. remanei* represent the most closely related gonochoristic/andro dioecious species pair (Cho et al., 2004; Kiontke et al., 2004), and thus make an especially informative comparative system for future research. More distantly related hermaphrodite species exist that have convergently evolved selfing (Fitch, 2002). The availability of both closely related species with different mating systems and multiple cases of parallel evolution of a complex trait makes soil nematodes an attractive system for developmental studies of reproductive adaptation. Rapid developments in genome sequencing and the advent of comparative genetics are making rigorous studies in these other species possible, and not only for sex determination. However, given the intrinsic organismal importance and remarkably rapid evolution of sex determination, it is likely that it will remain a major topic in nematode comparative biology for the foreseeable future.

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9. References

- Ahringer, J., and Kimble, J. (1991). Control of the sperm-oocyte switch in *Caenorhabditis elegans* hermaphrodites by the *fem-3* 3' untranslated region. *Nature* 349, 346–348. [Abstract Article](#)
- Ahringer, J., Rosenquist, T., Lawson, D., and Kimble, J. (1992). The *Caenorhabditis elegans* sex determining gene *fem-3* is regulated post-transcriptionally. *EMBO J.* 11, 2303–2310. [Abstract](#)
- Anderson, R. (2000). *Nematode parasites of vertebrates*, 2nd edn. (Wallingford, UK, CABI Publishing).
- Ashton, F.T., Bhopale, V.M., Holt, D., Smith, G., and Schad, G.A. (1998). Developmental switching in the parasitic nematode *Strongyloides stercoralis* is controlled by the ASF and ASI amphidial neurons. *J. Parasitol.* 84, 691–695. [Abstract](#)

- Barnes, T.M., and Hodgkin, J. (1996). The *tra-3* sex determination gene of *Caenorhabditis elegans* encodes a member of the calpain regulatory protease family. *EMBO J.* *15*, 4477–4484. [Abstract Article](#)
- Belfiore, M., Pugnale, P., Saudan, Z., and Puoti, A. (2004). Roles of the *C. elegans* cyclophilin-like protein MOG-6 in MEP-1 binding and germline fates. *Development* *131*, 2935–2945. [Abstract Article](#)
- Bull, J. (1981). Evolution of environmental sex determination from genotypic sex determination. *Heredity* *47*, 173–184.
- Bull, J. (1983). *Evolution of sex determining mechanisms*, Vol. 1 (Menlo Park, CA: Benjamin Cummings Publishing Co.).
- Carmi, I., Kopczynski, J.B., and Meyer, B.J. (1998). The nuclear hormone receptor SEX-1 is an X-chromosome signal that determines nematode sex. *Nature* *396*, 168–173. [Abstract Article](#)
- Chan, R.C., Severson, A.F., and Meyer, B.J. (2004). Condensin restructures chromosomes in preparation for meiotic divisions. *J. Cell Biol.* *167*, 613–625. [Abstract Article](#)
- Chen, P., Cho, S., Jin, S., and Ellis, R. (2001). Specification of germ cell fates by FOG-3 has been conserved during nematode evolution. *Genetics* *158*, 1513–1525. [Abstract](#)
- Chen, P., and Ellis, R.E. (2000). *tra-1A* regulates transcription of *fog-3*, which controls germ cell fate in *C. elegans*. *Development* *127*, 3119–3129. [Abstract](#)
- Chen, P., Singal, A., Kimble, J., and Ellis, R. (2000). A novel member of the tob family of proteins controls sexual fate in *Caenorhabditis elegans* germ cells. *Dev. Biol.* *217*, 77–90. [Abstract Article](#)
- Chitwood, B., and Chitwood, M. (1950). *Introduction to Nematology* (Baltimore: University Park Press).
- Cho, S., Jin, S.W., Cohen, A., and Ellis, R.E. (2004). A phylogeny of *caenorhabditis* reveals frequent loss of introns during nematode evolution. *Genome Res.* *14*, 1207–1220. [Abstract Article](#)
- Christie, J. (1929). Some observations on sex in the Mermithidae. *J. Exp. Zool.* *53*, 59–76. [Article](#)
- Clifford, R., Lee, M., Nayak, S., Ohmachi, M., Giorgini, F., and Schedl, T. (2000). FOG-2, a novel F-box-containing protein, associates with the GLD-1 RNA-binding protein and directs male sex determination in the *C. elegans* hermaphrodite germline. *Development* *127*, 5265–5276. [Abstract](#)
- Cline, T., and Meyer, B. (1996). Vive la difference: males vs. females in flies vs. worms. *Annu. Rev. Genet.* *30*, 637–702. [Abstract Article](#)
- Crittenden, S.L., Bernstein, D.S., Bachorik, J.L., Thompson, B.E., Gallegos, M., Petcherski, A.G., Moulder, G., Barstead, R., Wickens, M., and Kimble, J. (2002). A conserved RNA-binding protein controls germline stem cells in *Caenorhabditis elegans*. *Nature* *417*, 660–663. [Abstract Article](#)
- Dawes, H.E., Berlin, D.S., Lapidus, D.M., Nusbaum, C., Davis, T.L., and Meyer, B.J. (1999). Dosage compensation proteins targeted to X chromosomes by a determinant of hermaphrodite fate. *Science* *284*, 1800–1804. [Abstract Article](#)
- de Bono, M., and Hodgkin, J. (1996). Evolution of sex determination in *Caenorhabditis*: unusually high divergence of *tra-1* and its functional consequences. *Genetics* *144*, 587–595. [Abstract](#)
- Fitch, D., ed. (2002). Ch. 2 of Appendix, Phylogeny, of WormAtlas (<http://www.wormatlas.org>) (WormAtlas Consortium).
- Francis, R., Barton, M.K., Kimble, J., and Schedl, T. (1995). *gld-1*, a tumor suppressor gene required for oocyte development in *Caenorhabditis elegans*. *Genetics* *139*, 579–606. [Abstract](#)

- Gallegos, M., Ahringer, J., Crittenden, S., and Kimble, J. (1998). Repression by the 3'UTR of *fem-3*, a sex-determining gene, relies on a ubiquitous *mog*-dependent control in *Caenorhabditis elegans*. *EMBO J.* **17**, 6337–6347. [Abstract Article](#)
- Goodwin, E.B., Okkema, P.G., Evans, T.C., and Kimble, J. (1993). Translational regulation of *tra-2* by its 3' untranslated region controls sexual identity in *C. elegans*. *Cell* **75**, 329–339. [Abstract Article](#)
- Graham, P., Penn, J.K., and Schedl, P. (2003). Masters change, slaves remain. *BioEssays* **25**, 1–4. [Abstract Article](#)
- Haag, E., and Kimble, J. (2000). Regulatory elements required for development of *C. elegans* hermaphrodites are conserved in the *tra-2* homologue of *C. remanei*, a male/female sister species. *Genetics* **155**, 105–116. [Abstract](#)
- Haag, E., Wang, S., and Kimble, J. (2002). Rapid coevolution of the nematode sex-determining genes *fem-3* and *tra-2*. *Curr. Biol.* **12**, 2035–2041. [Abstract Article](#)
- Hagstrom, K.A., Holmes, V.F., Cozzarelli, N.R., and Meyer, B.J. (2002). *C. elegans* condensin promotes mitotic chromosome architecture, centromere organization, and sister chromatid segregation during mitosis and meiosis. *Genes Dev.* **16**, 729–742. [Abstract Article](#)
- Hamaoka, B.Y., Dann, C.E., III, Geisbrecht, B.V., and Leahy, D.J. (2004). Crystal structure of *Caenorhabditis elegans* *her-1* and characterization of the interaction between *her-1* and TRA-2A. *Proc. Natl. Acad. Sci. USA* **101**, 11673–11678. [Abstract Article](#)
- Hansen, D., Hubbard, E.J., and Schedl, T. (2004). Multi-pathway control of the proliferation versus meiotic development decision in the *Caenorhabditis elegans* germline. *Dev. Biol.* **268**, 342–357. [Abstract Article](#)
- Hansen, D., and Pilgrim, D. (1998). Molecular evolution of a sex determination protein. FEM-2 (pp2c) in *Caenorhabditis*. *Genetics* **149**, 1353–1362. [Abstract](#)
- Harlos, J., Brust, R., and Galloway, T. (1980). Observations on a nematode parasite of *Aedes vexans* (Diptera: Culicidae) in Manitoba. *Can. J. Zool.* **58**, 215–220.
- Hodgkin, J. (1986). Sex determination in the nematode *Caenorhabditis elegans*: analysis of *tra-3* suppressors and characterization of *fem* genes. *Genetics* **114**, 15–52. [Abstract](#)
- Hodgkin, J. (2002). Exploring the envelope. Systematic alteration in the sex-determination system of the nematode *Caenorhabditis elegans*. *Genetics* **162**, 767–780. [Abstract](#)
- Hodgkin, J., and Brenner, S. (1977). Mutations causing transformation of sexual phenotype in the nematode *Caenorhabditis elegans*. *Genetics* **86**, 275–287. [Abstract](#)
- Hodgkin, J., Zellan, J.D., and Albertson, D.G. (1994). Identification of a candidate primary sex determination locus, *fox-1*, on the X chromosome of *Caenorhabditis elegans*. *Development* **120**, 3681–3689. [Abstract](#)
- Honda, H. (1925). Experimental and cytological studies on bisexual and hermaphrodite free living nematodes, with special reference to problems of sex. *J. Morph. Physiol.* **40**, 191–233. [Article](#)
- Jan, E., Motzny, C.K., Graves, L.E., and Goodwin, E.B. (1999). The STAR protein, GLD-1, is a translational regulator of sexual identity in *Caenorhabditis elegans*. *EMBO J.* **18**, 258–269. [Abstract Article](#)
- Jan, E., Yoon, J.W., Walterhouse, D., Iannaccone, P., and Goodwin, E.B. (1997). Conservation of the *C. elegans* *tra-2* 3' UTR translational control. *EMBO J.* **16**, 6301–6313. [Abstract Article](#)
- Jin, S.W., Kimble, J., and Ellis, R.E. (2001). Regulation of cell fate in *Caenorhabditis elegans* by a novel cytoplasmic polyadenylation element binding protein. *Dev. Biol.* **229**, 537–553. [Abstract Article](#)
- Jones, A.R., Francis, R., and Schedl, T. (1996). GLD-1, a cytoplasmic protein essential for oocyte differentiation, shows stage- and sex-specific expression during *Caenorhabditis elegans* germline development. *Dev. Biol.* **180**, 165–183. [Abstract Article](#)

- Jones, A.R., and Schedl, T. (1995a). Mutations in *gld-1*, a femal germ cell-specific tumor suppressor gene in *Caenorhabditis elegans*, affect a conserved domain also found in Src-associated protein Sam68. *Genes Dev.* 9, 1491–1504. [Abstract](#)
- Kadyk, L.C., and Kimble, J. (1998). Genetic regulation of entry into meiosis in *Caenorhabditis elegans*. *Development* 125, 1803–1813. [Abstract](#)
- Kiontke, K., Gavin, N.P., Raynes, Y., Roehrig, C., Piano, F., and Fitch, D.H. (2004). *Caenorhabditis* phylogeny predicts convergence of hermaphroditism and extensive intron loss. *Proc. Natl. Acad. Sci. USA* 101, 9003–9008. [Abstract Article](#)
- Klein, R.D., and Meyer, B.J. (1993). Independent domains of the Sdc-3 protein control sex determination and dosage compensation in *C. elegans*. *Cell* 72, 349–364. [Abstract Article](#)
- Kraemer, B., Crittenden, S., Gallegos, M., Moulder, G., Barstead, R., Kimble, J., and Wickens, M. (1999). NANOS-3 and FBF proteins physically interact to control the sperm-oocyte switch in *Caenorhabditis elegans*. *Curr. Biol.* 9, 1009–1018. [Abstract Article](#)
- Kuwabara, P.E. (1996). Interspecies comparison reveals evolution of control regions in the nematode sex-determining gene *tra-2*. *Genetics* 144, 597–607. [Abstract](#)
- Kuwabara, P.E., and Kimble, J. (1995). A predicted membrane protein, TRA-2A, directs hermaphrodite development in *Caenorhabditis elegans*. *Development* 121, 2995–3004. [Abstract](#)
- Lamont, L.B., Crittenden, S.L., Bernstein, D., Wickens, M., and Kimble, J. (2004). FBF-1 and FBF-2 regulate the size of the mitotic region in the *C. elegans* germline. *Dev. Cell* 7, 697–707. [Abstract Article](#)
- Li, W., Boswell, R., and Wood, W. (2000). *mag-1*, a homolog of *Drosophila mago nashi*, regulates hermaphrodite germ-line sex determination in *Caenorhabditis*. *Dev. Biol.* 218, 172–182. [Abstract Article](#)
- Luitjens, C., Gallegos, M., Kraemer, B., Kimble, J., and Wickens, M. (2000). CPEB proteins control two key steps in spermatogenesis in *C. elegans*. *Genes Dev.* 14, 2596–2609. [Abstract Article](#)
- Lum, D., Kuwabara, P., Zarkower, D., and Spence, A. (2000). Direct protein-protein interaction between the intracellular domain of TRA-2 and the transcription factor *tra-1A* modulates feminizing activity in *C. elegans*. *Genes Dev.* 14, 3153–3165. [Abstract](#)
- Luz, J.G., Hassig, C.A., Pickle, C., Godzik, A., Meyer, B.J., and Wilson, I.A. (2003). XOL-1, primary determinant of sexual fate in *C. elegans*, is a GHMP kinase family member and a structural prototype for a class of developmental regulators. *Genes Dev.* 17, 977–990. [Abstract Article](#)
- Madl, J.E., and Herman, R.K. (1979). Polyploids and sex determination in *Caenorhabditis elegans*. *Genetics* 93, 393–402. [Abstract](#)
- Matsuda, M., Nagahama, Y., Shinomiya, A., Sato, T., Matsuda, C., Kobayashi, T., Morrey, C.E., Shibata, N., Asakawa, S., Shimizu, N., et al. (2002). DMY is a Y-specific DM-domain gene required for male development in the medaka fish. *Nature* 417, 559–563. [Abstract Article](#)
- Meyer, B.J. (2000). Sex in the worm: counting and compensating X-chromosome dose. *Trends Genet.* 16, 247–253. [Abstract Article](#)
- Miller, S.W., Hayward, D.C., Bunch, T.A., Miller, D.J., Ball, E.E., Bardwell, V.J., Zarkower, D., and Brower, D.L. (2003). A DM domain protein from a coral, *Acropora millepora*, homologous to proteins important for sex determination. *Evol. Dev.* 5, 251–258. [Abstract Article](#)
- Nayak, S., Goree, J., and Schedl, T. (2005). *fog-2* and the evolution of self-fertile hermaphroditism in *Caenorhabditis*. *PLoS Biology* 3, e6. [Abstract Article](#)

- Nigon, V. (1949). Les modalités de la reproduction et le déterminisme de sexe chez quelques Nematodes libres. Ann. Sci. Nat. Zool. ser. II, 2, 1–132.
- Nonet, M.L., and Meyer, B.J. (1991). Early aspects of *Caenorhabditis elegans* sex determination and dosage compensation are regulated by a zinc-finger protein. Nature 351, 65–68. [Abstract Article](#)
- Perry, M.D., Li, W., Trent, C., Robertson, B., Fire, A., Hageman, J.M., and Wood, W.B. (1993). Molecular characterization of the *her-1* gene suggests a direct role in cell signaling during *Caenorhabditis elegans* sex determination. Genes Dev. 7, 216–228. [Abstract](#)
- Petersen, J.J. (1985). Nematodes as biological control agents: Part I. Mermithidae. Adv. Parasitol. 24, 307–344. [Abstract](#)
- Pilgrim, D., McGregor, A., Jackle, P., Johnson, T., and Hansen, D. (1995). The *C. elegans* sex-determining gene *fem-2* encodes a putative protein phosphatase. Mol. Biol. Cell 6, 1159–1171. [Abstract](#)
- Pires-daSilva, A., and Sommer, R.J. (2004). Conservation of the global sex determination gene *tra-1* in distantly related nematodes. Genes Dev. 18, 1198–1208. [Abstract Article](#)
- Poinar, G., Jr. (2002). First fossil record of nematode parasitism of ants; a 40 million year tale. Parasitology 125, 457–459. [Abstract Article](#)
- Prahlad, V., Pilgrim, D., and Goodwin, E.B. (2003). Roles for mating and environment in *C. elegans* sex determination. Science 302, 1046–1049. [Abstract Article](#)
- Puoti, A., and Kimble, J. (1999). The *Caenorhabditis elegans* sex determination gene *mog-1* encodes a member of the DEAH-box protein family. Mol. Cell Biol. 19, 2189–2197. [Abstract](#)
- Puoti, A., and Kimble, J. (2000). The hermaphrodite sperm/oocyte switch requires the *Caenorhabditis elegans* homologs of PRP2 and PRP22. Proc. Natl. Acad. Sci. USA 97, 3276–3281. [Abstract Article](#)
- Raymond, C., Shamu, C., Shen, M., Seifert, K., Hirsch, B., Hodgkin, J., and Zarkower, D. (1998). Evidence for evolutionary conservation of sex-determining genes. Nature 391, 873–881. [Abstract Article](#)
- Sassaman, C. (1995). Sex determination and evolution of unisexuality in the *Conchostraca*. Hydrobiologia 298, 45–65. [Article](#)
- Sassaman, C., and Weeks, S.C. (1993). The genetic mechanism of sex determination in the conchostracean shrimp *Eulimnadia texana*. Am. Nat. 141, 314–328. [Article](#)
- Schedl, T., Graham, P.L., Barton, M.K., and Kimble, J. (1989). Analysis of the role of *tra-1* in germline sex determination in the nematode *Caenorhabditis elegans*. Genetics 123, 755–769. [Abstract](#)
- Spence, A., Coulson, A., and Hodgkin, J. (1990). The product of *fem-1*, a nematode sex-determining gene, contains a motif found in cell cycle control proteins and receptors for cell-cell interactions. Cell 60, 981–990. [Abstract Article](#)
- Stein, L., and others (2003). The genome sequence of *Caenorhabditis briggsae*: a platform for comparative genomics. PLoS Biology 1, 166–192. [Abstract Article](#)
- Stothard, P., Hansen, D., and Pilgrim, D. (2002). Evolution of the PP2C family in *Caenorhabditis*: rapid divergence of the sex-determining protein *FEM-2*. J. Mol. Evol. 54, 267–282. [Abstract Article](#)
- Streit, A., Li, W., Robertson, B., Schein, J., Kamal, I., Marra, M., and Wood, W. (1999). Homologs of the *Caenorhabditis elegans* masculinizing gene *her-1* in *C. briggsae* and the filarial parasite *Brugia malayi*. Genetics 152, 1573–1584. [Abstract](#)
- Wang, S., and Kimble, J. (2001). The *tra-1* transcription factor binds TRA-2 to regulate sexual fates in *Caenorhabditis elegans*. EMBO J. 20, 1363–1372. [Abstract Article](#)

Wilkins, A. (1995). Moving up the hierarchy: a hypothesis on the evolution of a genetic sex determination pathway. *BioEssays* 17, 71–77. [Abstract Article](#)

Yi, W., and Zarkower, D. (1999). Similarity of DNA binding and transcriptional regulation by *Caenorhabditis elegans* MAB-3 and *Drosophila melanogaster* DSX suggests conservation of sex determining mechanisms. *Development* 126, 873–881. [Abstract](#)

Zarkower, D., and Hodgkin, J. (1992). Molecular analysis of the *C. elegans* sex-determining gene *tra-1*: a gene encoding two zinc finger proteins. *Cell* 70, 237–249. [Abstract Article](#)

Zhang, B., Gallegos, M., Puoti, A., Durkin, A., Fields, S., Kimble, J., and Wickens, M.P. (1997). A conserved RNA binding protein that regulates sexual fates in the *C. elegans* hermaphrodite germ line. *Nature* 390, 477–484. [Abstract Article](#)



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